

实验记录

2018年7月24 9:11

date: 7.23

1. 50%的甘油进行分装，每管 500ul
2. 插枪头

date: 7.24

1. 倒 A 抗平板
2. Pet-21a 甘油菌平板划线活化，加 A 抗扩大培养。一共 3 管每管 10ml。1 管 NC 对照。
3. PUC57-UGT 引物设计
4. UGT 甘油菌活化

date: 7.25

- 1.
2. UGT 片段 PCR (PUC57backbone)

volume 50ul

| | |
|------------|----------|
| Master mix | 25 μ |
| F | 2 μ |
| R | 2 μ |
| Template | 4 μ |
| dd water | 17 μ |

program:

| | | | | | |
|------|-----|-------|-----|------|------|
| 98 | 98 | 52-60 | 72 | 72 | 4 |
| 2min | 10s | 15s | 15s | 5min | hold |

30 cycles

3. extract pET21a from E.coli
4. UGT carrying E.coli purification
5. pET-21a carrying E.coli purification

date: 7.26

1. UGT restriction endonuclease digest

| | |
|------------------|----------|
| Quick cut buffer | 10 μ |
| UGT pcr product | 20 μ |
| BamHI | 1 μ |
| EcoRI | 1 μ |
| dd water | 3 μ |
| pET-21a plasmid | 5 μ |

date: 7.27

1. extract PUC57 plasmid

date: 7.28

1. add EcoRI site on UGT fragment

volume 10 μ

| | |
|------------|-----------|
| Master mix | 5 μ |
| F | 0.4 μ |
| R | 0.4 μ |
| Template | 0.8 μ |
| dd water | 3.4 μ |

gradient PCR in 55-68°C. 8 columns total.

2. add EcoRI and NdeI site on UGT fragment

| | |
|------------|-----------|
| Master mix | 5 μ |
| F | 0.4 μ |
| R | 0.4 μ |
| Template | 0.8 μ |
| dd water | 3.4 μ |

gradient PCR in 55-65°C. 8 columns total.

4. PCR at best temperature

volume 100 μ

| | |
|------------|----------|
| Master mix | 50 μ |
| F | 4 μ |
| R | 4 μ |
| Template | 8 μ |
| dd water | 34 μ |

8 columns total. 4 single restriction endonuclease digest and 4 double restriction endonuclease digest

date: 7.30

1. single and double restriction endonuclease digest of UGT and pET21a

| | |
|------------------|----------|
| Quick cut buffer | 10 μ |
| UGT pcr product | 20 μ |
| BamHI | 1 μ |
| EcoRI | 1 μ |
| dd water | 3 μ |
| pET-21a plasmid | 5 μ |

2. UGT 与 pET-21a single and double restriction endonuclease digest and ligation

volume: 40 μ

| | |
|---------------------|----------|
| 10X ligation buffer | 4 μ |
| vector | 10 μ |
| Insert sequence | 20 μ |
| T4 DNA ligase | 2 μ |
| dd water | 4 μ |

date: 7.31

1. pET-21a plasmid extract. 4 column total.
2. pET-21a double restriction endonuclease digest verification
3. UGT-pET-21a EcoRI single restriction endonuclease digest:

| | |
|---------------------|----------|
| 10X ligation buffer | 4 μ |
| vector | 9 μ |
| Insert sequence | 25 μ |
| T4 DNA ligase | 2 μ |

4. UGT-pET-21a EcoRI/NdeI double restriction endonuclease digest:

| | |
|---------------------|----------|
| 10X ligation buffer | 4 μ |
| vector | 9 μ |
| Insert sequence | 25 μ |
| T4 DNA ligase | 2 μ |

date: 8.1

1. double restriction endonuclease digest of UGT fragment.

volume: 30 μ l

| | |
|-------------|----------|
| PCR product | 25 μ |
| EcoRI | 1 μ |
| NdeI | 1 μ |
| buffer | 3 μ |

pET-21a single restriction endonuclease digest

volume: 20 μ l

| | |
|----------|----------|
| pET-21a | 10 μ |
| EcoRI | 1 μ |
| NdeI | 1 μ |
| buffer | 2 μ |
| dd water | 6 μ |

2. UGT-pET-21a vector construction

| | |
|---------------------|----------|
| 10X ligation buffer | 4 μ |
| Vector | 5 μ |
| Insert fragment | 25 μ |
| T4 DNA ligase | 2 μ |

date: 8.2

1. UGT-pET-21a single restriction endonuclease digest and ligation

| | |
|---------------------|----------|
| 10X ligation buffer | 5 μ |
| Vector | 20 μ |
| Insert sequence | 20 μ |
| T4 DNA ligase | 2 μ |
| dd water | 3 μ |
| total | 50 μ |

2. Colony PCR

| | |
|----------|----------|
| supermix | 25 μ |
| F | 2 μ |
| R | 2 μ |
| Template | 4 μ |
| dd water | 17 μ |
| total | 50 μ |

5 columns total. One of them is negative control

3. Colony PCR of the double restriction endonuclease digest and ligation bacteria

| | |
|----------|----------|
| supermix | 25 μ |
| F | 2 μ |
| R | 2 μ |
| Template | 4 μ |
| dd water | 17 μ |
| total | 50 μ |

date: 8.3

1. Transfer the plasmids (single and double) into *E.coli*
2. Extract plasmids from the double restriction endonuclease digest and ligation transferred *E.coli*.
3. Transfer the third branch of ligation product
4. medium PCR

| | |
|----------|-----------|
| supermix | 5 μ |
| F | 0.4 μ |
| R | 0.4 μ |
| Template | 1 μ |
| dd water | 3.2 μ |

| | |
|-------|----------|
| total | 10 μ |
|-------|----------|

10 columns total

date: 8.5

1. Colony PCR of the UGT-transferred E.coli.

volume: 20 μ

| | |
|------------------|-----------|
| 2x super pcr mix | 10 μ |
| template | 1 μ |
| Primer1 | 0.8 μ |
| Primer2 | 0.8 μ |
| ddH2O | 7.4 μ |

The template is diluted into 1/10 concentration

PCR program:

| | | |
|------|------|---------|
| 98°C | 3min | |
| 98°C | 30s | 35cycle |
| 60°C | 30s | 35cycle |
| 72°C | 5s | 35cycle |
| 72°C | 2min | |
| 4°C | hold | |

date: 8.7

1. BL21 colony PCR

| | |
|----------|----------|
| supermix | 5 μ |
| F | 1 μ |
| R | 1 μ |
| Template | 1 μ |
| dd water | 2 μ |
| total | 10 μ |

10 column total. No lanes.

date: 8.8

1. single and double restriction endonuclease digest

| | |
|------------------|----------|
| Quick cut buffer | 3 μ |
| UGT pcr product | 25 μ |
| NdeI | 1 μ |
| EcoRI | 1 μ |
| pET-21a plasmid | 5 μ |
| total | 30 μ |

| | |
|------------------|----------|
| Quick cut buffer | 3 μ |
| UGT pcr product | 25 μ |
| EcoRI | 2 μ |
| pET-21a plasmid | 5 μ |
| total | 30 μ |

date: 8.9

1. redo UGT PCR

| | |
|-----------|----------|
| MCLAB mix | 20 μ |
| F | 4 μ |
| R | 4 μ |
| Template | 2 μ |
| dd water | 20 μ |
| total | 20 μ |

date: 8.10

1. restriction endonuclease digest and ligation then transfer into *E.coli*
2. pick 8 colonies then sequence
3. verify by PCR with redesign primers

| | |
|-----------|----------|
| super mix | 5 μ |
| F | 1 μ |
| R | 1 μ |
| Template | 1 μ |
| dd water | 2 μ |
| total | 10 μ |

8 columns total

result: 3 of them are correct plasmid. Amplify them for plasmid extracting.

date: 8.11

1. sequence the plasmid by Qingke company

date: 8.12

1. sequence the plasmid by Sangon Biotech, get same result of Qingke (correct).

date: 8.13

1. BL21transfer

date: 8.14

1. BL21 transferred strain PCR

positive. Store the strain in glycerol in -80 °C

2. Amplify the strain in 250ml LB medium.

date: 8.16

1. Lyse the bacteria by ultrasonic

2. Run the SDS-PAGE electrophoresis and stained with coomassie brilliant blue. The control group protein is too much so can get any useful information.